

Asian Journal of Environment & Ecology

Volume 23, Issue 10, Page 109-120, 2024; Article no.AJEE.124192 ISSN: 2456-690X

Evaluation of Phytoremediation Potential of Some Leguminous Crops on Crude Oil Polluted Soil in the Niger Delta Area

Achimota A. Dickson ^{a*}, Amadi Amadi ^b and Payou T Ogboin ^a

Department of Crop and Soil Science, Niger Delta University, Wilbe4rforce Island, Nigeria.
 Department of Microbiology, River State University, Port Harcourt, Nigeria.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: https://doi.org/10.9734/ajee/2024/v23i10614

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here:

https://www.sdiarticle5.com/review-history/124192

Original Research Article

Received: 23/07/2024 Accepted: 27/09/2024 Published: 03/10/2024

ABSTRACT

Since petroleum hydrocarbon contamination is a major pollution problem in the Niger Delta environment, the relative tolerance of some leguminous species to crude oil contamination and their potential in the phytoremediation of crude oil polluted soil was assessed in a greenhouse. Seeds of cowpea (*Vigna unguiculata L.* Walp), lablab (*Dolichos lablab*), mucuna (*Mucuna untilis*) and Soybean (*Glycine max*) were planted in soil polluted with 2 and 4% (v/w) crude oil and no pollution, 7 days after oil treatment in a 3x4 factorial arrangement, replicated thrice. Parameters assessed included germination percentage, plant height, leaf area, plant top and root biomass, .nodule number and plant nutrient N uptake. Whereas, at 4% pollution, germination of cowpea, lablab and

 $\hbox{*Corresponding author: E-mail: achimotadickson @ndu.edu.ng, achimotadickson @gmail.com;}$

Cite as: Dickson, Achimota A., Amadi Amadi, and Payou T Ogboin. 2024. "Evaluation of Phytoremediation Potential of Some Leguminous Crops on Crude Oil Polluted Soil in the Niger Delta Area". Asian Journal of Environment & Ecology 23 (10):109-20. https://doi.org/10.9734/ajee/2024/v23i10614.

soybean was depressed by 35, 40 and 60%, respectively, germination in mucuna, though delayed, was depressed by only 10%. Oil treatment significantly reduced (P<0.05) plant height, leaf area and biomass dry matter (DM) for all leguminous species except mucuna. In both oil polluted and unpolluted soils, mucuna formed more nodules and took up more soil N. At 4% pollution, mucuna produced more than 6, 4 and 3 times the biomass produced by soybean, lablab and cowpea, respectively. Therefore, at 4% pollution, if 1 ton dry matter of mucuna is turned-in as green manure, it may add 18kg/ha N to the soil compared to 11kg N for soybean. This study revealed the potentials of the legumes in the phytoremediation of crude oil contaminated soil and tolerance to oil pollution rating them in the order: mucuna>cowpea>lablab>soybean.

Keywords: Phytoremediation; mucuna; cowpea; lablab; soybean; oil pollution.

1. INTRODUCTION

Petroleum hydrocarbon contamination caused by crude oil spill is major problem in the Niger Delta environment [1] due to the occurrences of oil spills and the so much risks posed to human life and the environment [2,3]. This create several environmental problems including its adverse effect on soil condition [2,4,5,6] microorganisms and plants. As part of efforts to remedy the situation, investigators have examined the feasibility of some post-oil spill rehabilitation measures [4,7] on the rate of soil recovery and crop improvement. Three methods commonly used in treating petroleum hydrocarbon contamination chemical, are physical, and biological remediation, out of which phytoremediation. а biological remediation method, is becoming increasingly of interest, owing to its distinctive advantages as a green, safe technology with low cost and wide variety of potential applications [8]. But in most cases of oil pollution of soils, N and P alone have been observed to be most limiting to biodegradation and crop growth [9,7].

fertilizer the nutrients. nitrogen deficiencies have been observed to be most widespread in Nigeria [10]. And in the soils from the Coastal plain sand deposits of Southern Nigeria, Ijah et al. [11] reported that N is more limiting to crop production than P. The successes recorded by Amadi et al [7] when they used some N rich inorganic and organic nutrient supplements to reclaim an oil polluted soil is therefore understandable. Today, there is some level of awareness created among the people in the oil producing areas that application of N rich materials to polluted soils could accelerate soil recovery. But the fertilizer supplies from the Fertilizer Company National of Nigeria (NAFCON) cannot even meet the fertilizer requirements of the nation's farmers not to talk of using same for the reclamation of the several

polluted sites in the country. Since fertilizer is not readily available, the alternative could have been livestock, wastes but people show much aparthy to their use because of their offensive Phytoremediation odour and bulk. bioremediation are reported as cost-effective and friendly technologies environmentally Phytoremediation, an emerging technology, uses plants to clean up contaminants in environment [3] which is natural, environment friendly and more cost effective [8]. Several studies have consistently demonstrated that some plant species especially grasses and legumes have the potential to degrade petroleum hydrocarbons in soil. [13,14,3]. Since organic agriculture is taking the centre stage, there is need to source for alternative sources of N-rich materials that can be used for the rehabilitation of polluted sites. Using leguminous crops as green manure is worth trying because of their ability to fix N and enrich soils that are deficient in the nutrient. This study seeks to provide information on the relative tolerance of four leguminous crop species, cowpea (Vigna unguiculata L. Walp). Lablab (Dolichos lablab), Mucuna (Mucuna utilis) and Soybean (Glycine max), to oil pollution, and their potentials as green manure crops for reclaiming oil polluted sites in the country.

2. MATERIALS AND METHODS

2.1 Experimentation

The Nigerian Bonny light crude with a specific gravity of 0.8343 was used for this greenhouse studies. Surface soil (0 to 15cm depth) was collected from a one year fallow patch of land by the Acada village of the Rivers State University of Science and Technology, Port Harcourt, bulked, air dried and passed through a 2-mm sieve. Two and half kilograms of the composite sample was put into plastic planting buckets. The experiment was a 3 x 4 factorial fitted into a

completely randomized design (CRD), replicated thrice. The factors were oil (0, 2 and 4% (v/w) and the leguminous crops species (cowpea, lablab, mucuna and soybean). Details of the oil treatment levels are as shown in Table 1. Listed in Table 2 is the chemical characteristics of the sandy loam soil used for the studies.

All pots were watered consecutively for one week with tap water before the respective pots were polluted with the three levels of oil. One week after pollution, ten (10) seeds each of the leguminous species were planted separately to the respective pots thinned to 2 seedlings 2 weeks after planting (2WAP). The pots were watered at intervals of 2 days to about field capacity throughout the period of growth. Weeding was by hand picking and the hand-picked weeds deposited back in the respective pots to decompose. Growth parameters were monitored weekly and plants harvested 8WAP.

Germination counts were taken per pot per treatment combination and percentage germination calculated using the formula:

No of Seedlings (emerged) x 100 No. of Seedlings (planted)

Plant height was measured using a meter rule. Leaf area (cm²) was assessed by measuring the total length and breadth (at the broadest point) of the leaves on a plant and multiplying with a factor. At harvest, nodulation or nodule numbers

were taken after washing the roots in a sieve to prevent loss of the nodules. After washing and drying of plant tops and roots in an oven at 70°C for 3 days, plant top and root biomas weights were taken.

2.2 Soil Analyses

The soil samples were subjected to chemical analyses at the beginning of the experiment and after harvest of the crops. Standard methods were used in the analyses of the soils as described in Udo et al. [15] Estefan et al. [16].

2.3 Plant Tissue Analysis

Oven dried plant samples were ground in a hammer mill. Subsamples of the milled samples were digested using acid mixture (Perchloric acid – sulphuric acid – silicic acid mixture). After digestion, the digests were put in 100ml volumetric flasks and made to mark. Appropriate portions of the aliquots were then taken for nutrient concentration determination using standard methods.

2.4 Data Analysis

Simple linear regression analysis was carried out on the effect of oil treatment on germination percentage, plant height, leaf area, plant top biomass, plant root biomass, nodule number yield as well as soil properties and plant nutrient uptake.

Table 1. Treatment levels showing the volume of oil applied per 2.5kg soil and the leguminous species planted

Treatment	Rate of Application	Vol. of Oil	
Oil	0%	0ml	
	2%	50ml	
	4%	100ml	
Leguminous Species	Cowpea		
	Lablab		
	Mucuna		
	Soybean		

Table 2. Chemical characteristics of the experimental soil before treatment application

Texture	Sandy loam	
pH	4.77	
Ca ² +	2.80 cmol/kg	
Mg ² +	1.40 cmol/kg	
K+	0.14 cmol/kg	
PO ₄ ³⁻	82.63 mg/kg	
Org.C	1.59%	
Total N	0.08%	
C/N ratio	20	

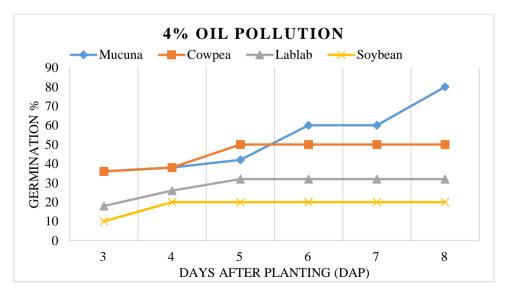
3. RESULTS AND DISCUSSION

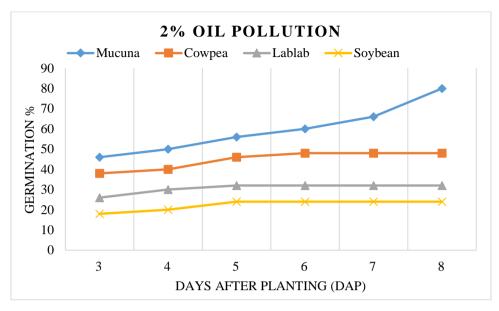
3.1 Effect of Oil Treatment on Seed Germination and Plant Growth Parameters

3.1.1 Seed germination

Germination counts (%) of the leguminous species as affected by oil contamination are illustrated in Fig. 1. Crude oil pollution at the 2% level delayed and decreased the germination of the leguminous species. For example, maximum germination occurred in all species for the unpolluted soil at 5DAP. But for the 2% oil level. it occurred at 6DAP and 8DAP for cowpea and mucuna, respectively. For lablab and soybean under the same level of pollution, germination stopped at 4DAP and 5DAP, respectively. At the 4% level also, germination of mucuna seeds were delayed to 8DAP while the germination of cowpea and lablab stopped at Germination of the soybean seeds stopped at 4DAP only. Comparing the rate of germination of the legume seeds, oil contamination reduced germination by 10, 35, 40 and 60% for mucuna, cowpea, lablab and soybean, respectively. Though germination in mucuna was delayed by oil pollution, 75% germination was recorded at both levels of pollution which is only 10% below the unpopulated condition (Fig. 1). A similar trend was reported by Lale et al. (2014). They recorded reduced plant height, laminar leaf area, number of leaves and shoot dry weight of cowpea plant cultivated in a soil contaminated by spent lubricating oil. Thus, mucuna is more

tolerant to the adverse effects of oil on germination relative to the other leguminous species. It might be possible that mucuna seed is of higher quality since germinability of seeds is a mere reflection of the quality of seed than environmental influence, with the environment exerting only a marginal influence on seedling establishment [7]. In this case, mucuna, whose seed sizes were relatively larger, probably had more stored energy for use by the developing seedlings. This might have enabled mucuna to withstand the adverse effect of oil germination. Our observation on the germination of these crops supports and re-enforced the findings of Uzoho et al., [17] Ekpo et al., [18] and Amadi et al. [7] on the effect of oil on These authors variously seed germination. observed that increasing the concentration of oil beyond 3% in soil reduced germination of maize and okra seeds. In this study, germination cowpea. lablab and sovbean seeds was decreased drastically by 2% oil in the soil. Hussien et al. [19] similarly, reported germination progressive decline in seed with increase crude percentage in percentage for V. rosea seeds in the soil. However, Adeyemi and Adeyemi [20] attributed the deleterious effects of crude oil on plant germination to cytotoxicity of crude constituents. The poor germination of the crops in this study might be attributed to the poor wettability and aeration in the soil, in addition to the toxic effects of the oil [21]. Also, oil coatings on seed surfaces affected the physiological functioning within the seed [7] leading to the poor germination.





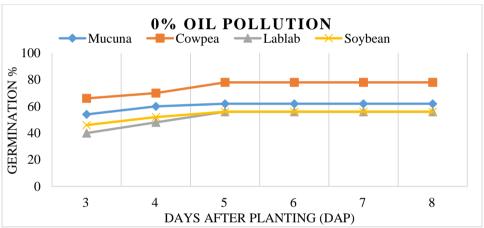


Fig. 1. Daily percentage germination of the leguminous species at the different pollution levels

3.2 Plant Growth

At 8WAP, oil pollution exerted significantly negative impact on all growth parameters except plant root biomass (Table 3). There was increase in height of all the legumes with time but increasing oil levels depressed heights of all species except mucuna and that of cowpea at 8WAP (Fig. 2). The height of mucuna at 8WAP varied from 130 cm at the 2% oil level to 140 cm at the 4% oil i.e. about 62% of the height of mucuna grown in unpolluted soil. Similarly, the heights of cowpea planted in soils polluted with 2% and 4% oil levels were 58% and 65% respectively, relative to that in the unpolluted soil. The heights of lablab and soybean on the other hand decreased with increase in oil concentration. Adeyemi and Adeyemi [20] reported decreasing plant height of Phaseolus vulgaris L with increasing oil level which was attributed to decreased availability of plant nutrients and oxidative stress.

depressing effect of increasing concentration on the growth of these legumes was also evident in the leaf area recorded for the crops except mucuna (Fig. 3). Like plant height, leaf area increased with time. The leaf area recorded for mucuna varied from 53cm2 in the 2% oil level to 59cm² in the 4% pollution level. These values correspond to 80% and 89% relative to that in the unpolluted soil. Using these two growth parameters to assess the crops, mucuna appears to be more tolerant to oil pollution. But Odeyemi and Immanuel (2023) reported reduction in leaf area and nodulation in the instance of crude oil in soil, and ascribed it to impaired metabolic processes in plants growing in the polluted soil. The results in this study might be as a result of the ability of mucuna to form

more nodules even in polluted condition (Fig. 4). Whereas mucuna formed 82, 48 and 23 nodules at 0, 2 and 4% oil levels, that of lablab was only 37, 12 and 13, respectively. It is possible that the greater number of nodules in mucuna enhanced more N uptake which encouraged more luxuriant growth of the crop both in polluted and unpolluted conditions. Moreover, mucuna seems to be better in withstanding oxidative stress and impaired metabolic processes in oil polluted soils.

3.3 Effect of Oil Treatment on Yield

3.3.1 Dry matter yield

As already recorded for the growth parameters, whereas, increasing concentration of depressed plant top and root biomass of cowpea, lablab and soybean, that of mucuna decreased from the unpolluted condition to the 2% oil level and increased when the oil level increased to 4% The result tend to corroborate the (Fia. 5). results of growth parameters that mucuna is more tolerant to oil pollution among the 4 leguminous species. This perhaps, is due to its ability to nodulate well in polluted soils. At 4% oil level, mucuna was able to produce 56% and 86% plant top and root biomass relative to that in Lablab on the other hand unpolluted soil. produced only 24% and 32% of the respective biomass relative to that in polluted condition. Pre-planting oil pollution, perhaps, exerted deleterious effect on the performance of these legumes by altering soil physical, chemical and biological properties [2,7]. These in turn affected the rate of synthesis and translocation of vital mineral nutrients in the plants. Mucuna probably have greater ability to withstand the deleterious effects.

3.4 Effect of Oil Treatment on Soil Properties

3.4.1 Soil properties

The results of the effect of oil pollution and the leguminous species on soil chemical properties

are shown on Table 4. Soil pH values increased slightly with increasing levels of oil. results agreed with the findings of Macci et al. [6] and Uzoho et al. [17] who variously reported that oil buffered acid soil pHs toward neutral values. This shift in pH is understandable because the sandy loam soil used is low in organic matter and therefore low in buffering characteristics. Unlike pH, exchangeable cations were not affected appreciably. Some decrease in soil P due to oil pollution was however noticed. Organic C increased with increase in oil level: the increase is more marked than pH. This increase might be due to addition of carbon from the degradation of the hydrocarbons to the organic C pool [6]. On the other hand, N did not vary significantly. Rather, the correlation between oil concentration and soil N was positive (Table 5). But Amadi et al. [7] working on a similar soil reported largescale reduction in soil N due to oil treatment which they attributed to N immobilization by microbial biomass and plant tissues. crude oil is generally low in N that is accessible to microbes [22] and hydrocarbon degrading organisms naturally, are present in any soil whose population size increases rapidly in response to input of oil into the environment [23] there must have been an increase in the population of microorganisms, especially the hydrocarbon degrading forms in the oil polluted pots. Ordinarily, these organisms require N and other metabolic feed stocks to degrade the oil which results in the reduction of N and P in oil polluted soils [21,18]. But the results in Table 6 indicated no reduction in N of the polluted soils over the unpolluted. This may mean that the Nfixing organisms associated with the legumes helped to fix and increase the N level of the polluted soils. Ekpo et al. [18] and Uzoho et al. [17] had earlier reported slight increases in soil N due to increases in oil doses which they attributed to increased activity of N-fixing organisms in the oil-contaminated soils. These emphasized the role results played by leguminous species in fixing and increasing N level even in oil polluted soils.

Table 3. The relationship between oil treatment and percentage germination (8DAP), plant height (8WAP), leaf area (8WAP), nodule number, plant top and root biomass production at harvest as expressed by correlation coefficient (r value) and regression equation

Correlation Factor	Significance (r) level	Regression Equation
Oil: Germination	-0.655 ***	Y = 51.26-0.05X
Oil: Plant Height	-0.353*	Y = 64.13-0.01 X
Oil: Leaf Area	-0.457**	Y = 42.02 - 0.05 X
Oil: Plant Top Biomass	-0.608*	Y = 7.49 - 0.22X
Oil: Plant Root Biomass	-0.172ns	Y = 3.18 - 0.11 X
Oil: Nodule Number	-0.740**	Y = 32.45 - 0.06 X

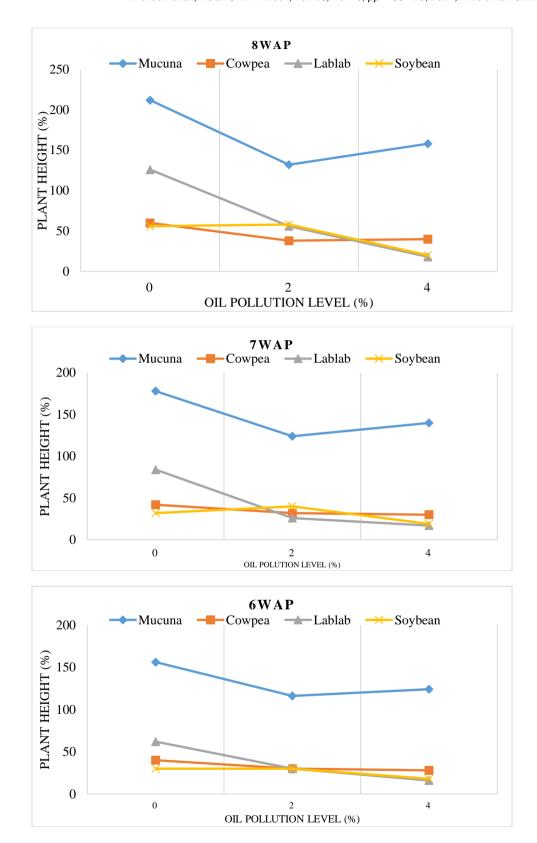
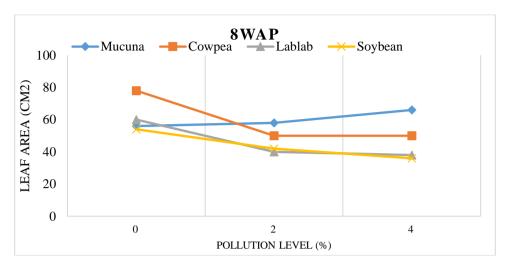
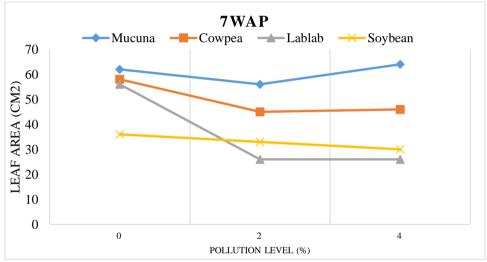


Fig. 2. Mean plant height (Weekly) of the leguminous species at the three oil levels





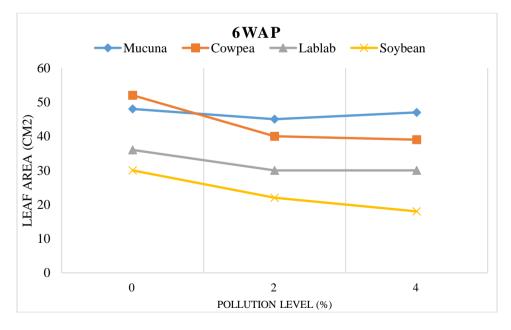


Fig. 3. Mean leaf (Weekly) of the leguminous species at the three oil levels

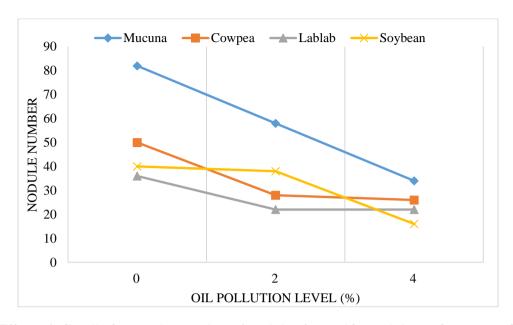


Fig. 4. Effect of oil pollution on the number of nodules formed in each leguminous species

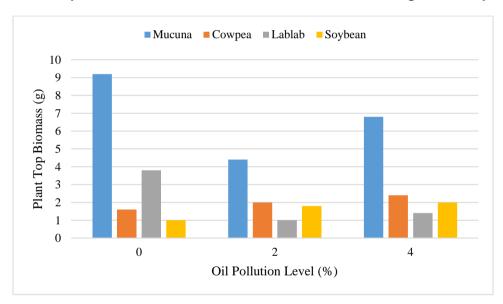


Fig. 5. Effect of crude oil pollution (%) on biomass production of the legumes

The results on Table 5 show that oil pollution had significant positive correlations with soil pH, organic C and C/N ratio. This positive relationship is an indication that their levels in the soil were increased by oil treatment.

3.5 Effect of Oil Treatment on Plant Nutrient Uptake

3.5.1 Nutrient uptake

The concentration of nutrients in the leguminous crops at the different oil levels after harvest are as presented in Table 6. And in Table 7 is

shown the correlation between oil treatment and the nutrients in the plants. Oil treatment had a significantly negative correlation with P in the crops (r=-0.769*). The negative correlation between oil treatment and plant nutrient concentration is an indication that their uptake by these crops was adversely affected. Apart from Mg and K, all the others gave negative correlation with oil treatment.

From the results in Table 6, the highest concentration of N in plants harvested from soils polluted with 2% oil was lablab (1.78%) followed by mucuna (1.76%) while the least was in

soybean (1.02%). At 4% pollution level, mucuna with 1.76% concentration was highest while soybean was least (1.11%). Thus, 1 ton of mucuna dry matter from 4% oil polluted field, turned-in as green manure could add about 18kg of N to the soil while soybean under similar condition will return only 11kg N. The total biomass production (plant top and root biomass) at the 4% oil pollution level for

mucuna was 17.3g against 2.8g for soybean (Fig. 5). These results revealed that mucuna produced more than 6, 4 and 3 times the biomass produced by soybean, lablab and cowpea, respectively. Combining the biomass production and nutrient uptake qualities for these leguminous species, the crops maybe rated in the order mucuna> cowpea> lablab>soybean.

Table 4. Effect of oil treatment on soil chemical properties planted to the different leguminous species

Oil Treatment %	Leguminous Species	pH (H ₂ O) 1:25	Org. C %	Total N %	Ca++	Mg++	K+	Avail P mg/kg	C/N ratio
0	Cowpea	4.66	1.79	0.08	2.40	0.62	0.12	110.52	22
	Lablab	4.69	1.76	0.07	2.00	0.29	0.16	105.26	25
	Mucuna	4.83	1.40	0.06	2.20	0.81	0.15	92.98	23
	Soybean	4.78	1.68	0.07	2.60	0.41	0.10	66.66	24
2	Cowpea	4.85	4.95	0.09	2.40	0.40	0.17	52.63	55
	Lablab	4.88	3.35	0.11	2.40	0.31	0.18	98.24	30
	Mucuna	4.68	4.68	0.06	2.20	0.60	0.14	78.94	78
	Soybean	4.76	4.74	0.07	2.60	0.22	0.13	43.86	68
4	Cowpea	5.06	4.55	0.08	3.00	0.28	0.18	43.86	57
	Lablab	4.99	5.88	0.11	2.80	0.61	0.17	87.72	53
	Mucuna	4.92	4.99	0.07	2.20	0.80	0.15	75.44	71
	Soybean	4.89	5.54	0.09	2.40	0.41	0.14	43.86	62

Table 5. The relationship between oil treatment and soil properties after harvest as expressed by correlation coefficient (r value) and regression equation

Correlation Factor	Significance (r) level	Regression Equation
Oil Treatment Vs		
рH	+0.769*	Y = -16.23 + 10.53 X
Ca2+	+0.456ns	Y = -3.11 + 2.77 X
Mg2+	+0.016ns	Y = 0.74 - 0.13 X
K+	+0.475ns	Y = 65.61 + 32.88 X
Org. C	+0.910***	Y = 1.94 + 0.92 X
N	+0.438ns	Y =- 87.42 + 43.75 X
Р	-0.538ns	Y = -75.08 - 0.04 X
C/N ratio	+0.752**	Y = 47.21 + 0.06 X

Table 6. Effect of oil treatment on nutrient concentration in the tissues of the legumes

Oil Treatment	Leguminous Species	N	Р	%K	Ca	Mg
0	Cowpea	1.43	0.53	4.00	1.74	2.67
	Lablab	1.82	0.26	2.75	1.80	4.96
	Mucuna	1.46	0.39	2.50	1.46	2.11
	Soybean	1.32	0.25	2.88	1.82	2.86
2	Cowpea	1.39	0.08	4.88	1.43	2.95
	Lablab	1.78	0.05	4.00	1.26	3.47
	Mucuna	1.76	0.05	2.75	1.35	4.47
	Soybean	1.02	0.06	3.75	1.35	3.02
4	Cowpea	1.32	0.05	4.38	1.28	3.22
	Lablab	1.53	0.06	3.50	1.58	5.76
	Mucuna	1.76	0.04	2.63	1.88	4.08
	Soybean	1.11	0.05	2.75	1.20	2.82

Table 7. The relationship between oil treatment and nutrient concentration in the leguminous species as expressed by correlation coefficient (r) value and regression equation

Correlation Factor	Significance (r) level	Regression Equation
Oil Treatment N	-0.044ns	Y = 2.04 - 0.28 X
Р	0.798**	Y = 17.82 - 8.83 X
K	+0.152ns	Y = 2.74 + 0.33X
Ca	-0.387ns	Y = 6.95 - 2.72 X
Mg	+0.327ns	Y = 2.50 + 0.52 X

4. CONCLUSION

And whereas increasing oil concentration increased pH and organic matter levels in the soil, it had no noticeable effect on soil N. This is because the N-fixing organisms and their associated legumes helped to fix and improve N level even in oil polluted soil condition. results showed that whereas oil treatment decreased germination in cowpea, lablab and soybean, germination in mucuna was only delayed and not decreased even at the 4% oil level. The same trend was observed in plant height, leaf area and biomass production. Mucuna in polluted soils formed more nodules than others and mobilized more soil N into its These findings point to the fact that tissues. among the legumes studied, mucuna has the greatest potential for use as green manure and phytoremediation of crude oil polluted soils. It is however recommended that further study should be carried out with higher oil concentrations supplemented with organic manures.

DISCLAIMER (ARTIFICIAL INTELLIGENCE)

Author(s) hereby declare that NO generative Al technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of this manuscript.

ACKNOWLEDGEMENTS

The authors acknowledge the support of the technical staff of the Department of Crop and Soil Science Laboratory, Niger Delta University, Wilberforce Island, Nigeria in analyzing the samples.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Oyedeji AA. Immanuel OM. Phytoremediation Potential of Senna fistula

- L. in the Decontamination of an Oil-Polluted Soil. J. Appl. Sci. Environ. Manage. 2023;27(12):2967-2972. DOI:https://dx.doi.org/10.4314/jasem.v27i1 2.39
- 2. Nemati B. Baneshi MB. Akbari Rouhullah Dehghani R. Mostafaii G. Phytoremediation of pollutants in oil-contaminated soils by Alhagi camelorum: an evaluation and modeling. Scientific Reports; 2024. Available:www.nature.com/scientificreports / 12p Available:https://doi.org/10.1038/s41598-024-56214-v
- 3. Ayibanoa Ibaba and Achimota Dickson. Organic manure enhanced phytoremediation of crude oil contaminated soil in the Niger Delta: the potentials of cowpea (*Vigna unguiculata*). Global Journal of Agricultural Research. 2024;12 (10):16-24.
- 4. Ossai IC, Ahmed A, Hassan A, Hamid FS. Remediation of soil and water contaminated with petroleum hydrocarbon: A review. Environmental Technology & Innovation. 2020;17:100526.
- 5. Hentati O, Lachhab R, Ayadi M, Ksibi M. **Toxicity** assessment petroleumfor terrestrial contaminated soil using invertebrates and plant bioassays. Environmental monitoring and assessment. 2013;185:2989-2998.
- 6. Macci C, Doni S, Peruzzi E, Bardella S, Filippis G, Ceccanti B, Masciandaro G. A real-scale soil phytoremediation. Biodegradation. 2013;24:521-538.
- 7. Amadi A, Aickson AA, Maate GO. Remediation of oil polluted soils: I. Effect of Organic and inorganic nutrient supplements on the performance of Maize (*Zea Mays* L.). Water, Air and Soil Pollution. 1993;66:59-76
- 8. Xiao N, Liu R, Jin C, Dai Y. Efficiency of five ornamental plant species in the phytoremediation of polycyclic aromatic

- hydrocarbon(PAH)-contaminated soil. Ecol Eng. 2015;75:384–391.
- Available:whttps:// doi. org/ 10. 1016/j. ecole ng. 2014. 12. 008
- 9. Chijioke-Osuji CC, Ibegbulam-Njoku PN, Belford EJ. Biodegradation of crude oil polluted soil by Co-composting with agricultural wastes and inorganic fertilizer. Biodegradation. 2014;4(6).
- Kamara AY, Menkir A, Ajala SO, Kureh I. Performance of diverse maize genotypes under nitrogen deficiency in the northern Guinea savanna of Nigeria. Experimental Agriculture. 2005;41(2):199-212.
- 11. Ijah CJ, Umoh FO, Essien GG, Edem TT. Phosphorus Status of Coastal Plain Sands of Akwa Ibom State, Nigeria; 2021.
- 12. Hussein SZ, Ahmad K, Hegazy NHM, Mohamed AE, Shafik DI, Gehan S. Eco-physiological response and genotoxicity-induced by crude petroleum oil in the potential phytoremediator *Vinca rosea* L.; 2022. Available:whttps://doi.org/10.1186/s43141-022-00412-6
- 13. Godwin U, Akpan, Peter E, Usuah. Phytoremediation of Diesel Oil Polluted Soil by Fluted Pumpkin (*Telfairia Occidentalis Hook* F.) in Uyo, Niger Delta Region, Nigeria Journal of Environment and Earth Science; 2014.
- Akpokodje OI, Uguru H, Esegbuyota D. Evaluation of phytoremediation potentials of different plants' varieties in petroleum products polluted soil. Global Journal of Earth and Environmental Science. 2019; A47C33942
- 15. Udo EJ, Ibia TO, Ogunwale JA, Ano AO, Esu IE. Manual of Soil, Plant and Water Analyses. Sibon Books Ltd., Lagos. 2009;188.

- Estefan G, Sommer R, Ryan J. Methods of Soil, Plant and Water Analysis: A manual for the West Asia and North Africa., International Center for Agricultural Research in the Dry Areas (ICARDA); 2013.
- Uzoho B, Oti NN, Onweremadu EU. Effect of crude oil pollution on maize growth and soil properties in Ihiagwa, Imo State, Nigeria. International Journal of Agriculture and Rural Development. 2004;5(1):91-100.
- 18. Ekpo IA, Agbor RB, Okpako EC, Ekanem EB. Effect of crude oil polluted soil on germination and growth of soybean (*Glycine max*); 2012.
- Hussein SZ, Nashwa H, Ahmad KH, Mohamed AE, Nermen H, Gehan S. Phytoremediation of Crude Petroleum Oil Pollution: A Review. Egyptian Journal of Botany. 2022;62(3):611-640.
- 20. Adeyemi O, Adeyemi O. Effect of Crude Oil Contaminated Soil on Phaseolus vulgaris L. World J. Inter. Res. 2020;8(2): 28-33.
- 21. Kekere O, Ikhajiagbe B, Apela BR. Effects of crude petroleum oil on the germination, growth and yield of *Vigna unguiculata Walp* L. Journal of Agriculture and Biological Sciences. 2011;2(6):158-165.
- 22. Wolicka D, Borkowski A. Microorganisms and crude oil. Introduction to enhanced oil recovery (EOR) processes and bioremediation of oil-contaminated sites. 2012;113-142.
- Vincent AO, Felix E, Weltime MO, Izeiyamu OK, Daniel EE. Microbial degradation and its kinetics on crude oil polluted soil. Research Journal of Chemical Sciences; 2011.
 ISSN, 2231, 606X.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of the publisher and/or the editor(s). This publisher and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.

© Copyright (2024): Author(s). The licensee is the journal publisher. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/124192